Product sheet

P3X63Ag8 | 305171

General information

Description	This cell ine is derived from the P3K27 cell line (a tissue culture line from the MOPC-21 plasmacytoma in BALB/c mice). The cells are resistant to 8-azaguanine (20 µg/mL) but sensitive to HAT. Due to a deficiency in 3-ketosteroid reductase activity, the cells have been reported to be cholesterol auxotrophs. This cell line was tested for being not infected by ectromelia virus (mousepox).
Organism	Mouse
Tissue	B Lymphocyte, Phlogocyte, Myeloma
Synonyms	P3x63Ag8, P3-x63-Ag8, P3/x63-Ag8, P3/x63 Ag8, P3/x63/Ag8, P3-x63Ag8, P3x63 Ag8, P3x63 Ag8, P3 x 63Ag8, P3 x 63 Ag8, x63-Ag8, x63-AG8, x63-Ag8, P3x63, x63, GM03571

Characteristics

Gender	Female
Morphology	Lymphoblast
Growth properties	Suspension

Identifiers / Biosafety / Citation

Citation	P3x63Ag8 (Cytion catalog number 305171)
Biosafety level	1

Expression / Mutation

Protein expression	Immunoglobulin, monoclonal antibody
Antigen expression	H-2d

Handling

Culture Medium	DMEM

Medium supplements	10% FBS, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 1.5 g/L NaHCO3, w: 1.0 mM Sodium pyruvate
Passaging solution	The utilization of a passaging solution is not necessary when passaging cells that are cultured in suspension. The appropriate procedure is to dilute the cells in accordance with the indicated guidelines.
Doubling time	16 to 26 hours
Subculturing	Resuspend cell suspension in the flask and take representative aliquote to count the cell number per ml. Dilute cell suspension to 1 x 10^5 cells/ml with fresh medium and transfer into new flasks.
Split ratio	2×10^5 cells/mL
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)
Handling of cryopreserved cultures	The cells come deep-frozen shipped on dry ice. Please make sure that the vial is still frozen. If immediate culturing is not intended, the cryovial must be stored below -150 degree Celsius after arrival. If immediate culturing is intended, please follow the below instructions: Quickly thaw by rapid agitation in a 37 degree Celsius water bath within 40-60 seconds. The water bath should have clean water containing an antimicrobial agent. As soon as the sample has thawed, remove the cryovial from the water bath. A small ice clump should still remain and the vial should still be cold. From now on, all operations should be carried out under aseptic conditions. Transfer the cryovial to a sterile flow cabinet and wipe with 70% alcohol. Carefully open the vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of culture medium (room temperature). Resuspend the cells carefully. Centrifuge at 300 x g for 3 min and discard the supernatant. The centrifugation step may be omitted, but in this case the remains of the freeze medium have to be removed 24 hours later. Resuspend the cells carefully in 10 ml fresh cell culture medium and transfer them into two T25 cell culture flasks. All further steps are described in the subculture section.
Handling of proliferating cultures	One or two cell culture flasks come filled with cell culture medium. Collect the entire medium in 1 or 2 x 50 ml centrifuge tubes, respectively. Carefully add 5 ml of cell culture medium to each T25 cell culture flask. Control the cell morphology and confluency under the microscope. Incubate at 37 degree Celsius for a minimum of 24 hours. Spin down the collected medium at 300 x g for 3 minutes to collect the cells which may have detached during transit. If a cell pellet is visible, resuspend the cells in 5 ml of cell culture medium and transfer to a T25 cell culture flask. Incubate at 37 degree Celsius for a minimum of 24 hours.

Quality control / Genetic profile / HLA

Sterility	Mycoplasma contaminations are ruled out through PCR-based and luminescence-based mycoplasma assays. The absence of bacterial, fungal or yeast contamination is controlled through daily visual cell monitoring.