Product sheet

RPMI 2650 | 300323

General information

Description	The cells are positive for keratin by immunoperoxidase staining.
Organism	Human
Tissue	Nasal septum
Disease	Squamous cell carcinoma
Metastatic site	Pleural effusion
Synonyms	RPMI-2650, RPMI2650, Roswell Park Memorial Institute 2650

Characteristics

Age	52 years
Gender	Male
Ethnicity	Caucasian
Morphology	Epithelial-like
Growth properties	Monolayer, adherent

Identifiers / Biosafety / Citation

Citation	RPMI 2650 (Cytion catalog number 300323)
Biosafety level	1

Expression / Mutation

Isoenzymes	G6PD, B
Reverse transcriptase	Negative

Products	Mucoid, keratin

Handling

Culture Medium	EMEM
Medium supplements	10% FBS, w: 2 mM L-Glutamine, w: 1.5 g/L NaHCO3, w: EBSS, w: 1 mM Sodium pyruvate, w: NEAA
Passaging solution	Accutase
Doubling time	48 hours
Subculturing	Seed at 2 x10^4 cells/cm2 during regular cell culture
Split ratio	A ratio of 1:2 to 1:4 is recommended
Fluid renewal	2 times per week
Freezing recovery	After thawing allow the cells to recover from the freezing process for at least 24 to 48 hours.
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)
Handling of cryopreserved cultures	The cells come deep-frozen shipped on dry ice. Please make sure that the vial is still frozen. If immediate culturing is not intended, the cryovial must be stored below -150 degree Celsius after arrival. If immediate culturing is intended, please follow the below instructions: Quickly thaw by rapid agitation in a 37 degree Celsius water bath within 40-60 seconds. The water bath should have clean water containing an antimicrobial agent. As soon as the sample has thawed, remove the cryovial from the water bath. A small ice clump should still remain and the vial should still be cold. From now on, all operations should be carried out under aseptic conditions. Transfer the cryovial to a sterile flow cabinet and wipe with 70% alcohol. Carefully open the vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of culture medium (room temperature). Resuspend the cells carefully. Centrifuge at 300 x g for 3 min and discard the supernatant. The centrifugation step may be omitted, but in this case the remains of the freeze medium have to be removed 24 hours later. Resuspend the cells carefully in 10 ml fresh cell culture medium and transfer them into two T25 cell culture flasks. All further steps are described in the subculture section.

Handling of proliferating cultures	One or two cell culture flasks come filled with cell culture medium. Collect the entire medium in 1 or 2 x 50 ml centrifuge tubes, respectively. Carefully add 5 ml of cell culture medium to each T25 cell culture flask. Control the cell morphology and confluency under the microscope. Incubate at 37 degree Celsius for a minimum of 24 hours. Spin down the collected medium at 300 x g for 3 minutes to collect the cells which may have detached during transit. If a cell pellet is visible, resuspend the cells in 5 ml of cell culture medium and transfer to a T25 cell culture flask. Incubate at 37 degree Celsius for a minimum of 24 hours.

Quality control / Genetic profile / HLA

Sterility	Mycoplasma contaminations are ruled out through PCR-based and luminescence-based mycoplasma assays. The absence of bacterial, fungal or yeast contamination is controlled through daily visual cell monitoring.
STR profile	**Amelogenin**: x,x **CSF1PO**: 9,11 **D13S317**: 11,12 **D16S539**: 11,12 **D5S818**: 12,13 **D7S820**: 8,11 **TH01**: 6,8 **TPOX**: 8 **vWA**: 16,18 **D3S1358**: 17 **D21S11**: 28,33.2 **D18S51**: 16 **Penta E**: 11,19 **Penta D**: 9,10 **D8S1179**: 9,13 **FGA**: 23,25
HLA alleles	**A***: 02:01:01, 03:01:01 **B***: 07:02:01, 35:01:01 **C***: 03:03:01, 07:02:01 **DRB1***: 07:01:01, 08:01:01G **DQA1***: 02:01:01, 04:01:01 **DQB1***: 03:03:02, 04:02:01 **DPB1***: 04:01:01 **E**: 01:01:01, 01:03