Product sheet

P388-D1 | 400308

General information

Description	A subclone of this line [P388 D1(IL-1)] produces high levels of interleukin-1 (IL-1).
Organism	Mouse
Tissue	Hematopoietic
Disease	Lymphoid neoplasma
Synonyms	P-388D1, P388D1, P388.D1, P3 88 D1

Characteristics

Gender	Female
Morphology	Round cells
Cell type	Macrophage
Growth properties	Suspension

Identifiers / Biosafety / Citation

Citation	P388-D1 (Cytion catalog number 400308)
Biosafety level	1

Expression / Mutation

Antigen expression	H-2d
Tumorigenic	Yes, in nude mice
Viruses	MAP-test negative: Sendai, Ektromelie (mousepox), Polyoma, K-Virus, Kilham, Reo 3, PVM, LCM, M.pulmonis, MVM, Theiler's GD VII, Toolan's H-1, MHV, LDV, RCV/SDA, M-Adenovirus, B.piliformis.
Reverse transcriptase	Positive

Handling

Culture Medium	RPMI 1640
Medium supplements	10% FBS, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3
Passaging solution	The utilization of a passaging solution is not necessary when passaging cells that are cultured in suspension. The appropriate procedure is to dilute the cells in accordance with the indicated guidelines.
Subculturing	Cultures can be maintained by addition of fresh medium or replacement of medium after centrifugation. The optimum cell density is at about 6 x 10^5 cells/ml.
Seeding density	Subculture at 1 x 10^6 viable cells/ml
Fluid renewal	Every 2 days
Freezing recovery	Fast. Allow the cells to recover from the freezing process for 24 hours. Then count the cells and dilute if > 10^6 viable cells are present.
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)
Handling of cryopreserved cultures	The cells come deep-frozen shipped on dry ice. Please make sure that the vial is still frozen. If immediate culturing is not intended, the cryovial must be stored below -150 degree Celsius after arrival. If immediate culturing is intended, please follow the below instructions: Quickly thaw by rapid agitation in a 37 degree Celsius water bath within 40-60 seconds. The water bath should have clean water containing an antimicrobial agent. As soon as the sample has thawed, remove the cryovial from the water bath. A small ice clump should still remain and the vial should still be cold. From now on, all operations should be carried out under aseptic conditions. Transfer the cryovial to a sterile flow cabinet and wipe with 70% alcohol. Carefully open the vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of culture medium (room temperature). Resuspend the cells carefully. Centrifuge at 300 x g for 3 min and discard the supernatant. The centrifugation step may be omitted, but in this case the remains of the freeze medium have to be removed 24 hours later. Resuspend the cells carefully in 10 ml fresh cell culture medium and transfer them into two T25 cell culture flasks. All further steps are described in the subculture section.
Handling of proliferating cultures	One or two cell culture flasks come filled with cell culture medium. Collect the entire medium in 1 or 2 x 50 ml centrifuge tubes, respectively. Carefully add 5 ml of cell culture medium to each T25 cell culture flask. Control the cell morphology and confluency under the microscope. Incubate at 37 degree Celsius for a minimum of 24 hours. Spin down the collected medium at 300 x g for 3 minutes to collect the cells which may have detached during transit. If a cell pellet is visible, resuspend the cells in 5 ml of cell culture medium and transfer to a T25 cell culture flask. Incubate at 37 degree Celsius for a minimum of 24 hours.

Quality control / Genetic profile / HLA

Sterility	Mycoplasma contaminations are ruled out through PCR-based and luminescence-based mycoplasma assays. The absence of bacterial, fungal or yeast contamination is controlled through daily visual cell monitoring.